

# Interaction of novel Dobzhansky–Muller type genes for the induction of hybrid lethality between *Gossypium hirsutum* and *G. barbadense* cv. Coastland R4-4

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**Abstract** Hybrid lethality was identified in interspecific hybrids between two cotton species, *Gossypium hirsutum* and *G. barbadense* cv. Coastland R4-4 (R4-4). Genetic analysis indicated that the lethal symptom was controlled by two dominant complementary genes, one from *G. hirsutum* and another from R4-4. Microsatellite mapping identified the location of the causal gene in *G. hirsutum* as chromosome D8, while the R4-4 gene was placed on chromosome D11. Our data indicate that these genes conform to the Dobzhansky–Muller model, and are novel for the induction of hybrid lethality in *Gossypium*. Following the genetic nomenclature, we propose that the two novel Dobzhansky–Muller genes from *G. hirsutum* and from R4-4 be named *Le*<sub>3</sub> and *Le*<sub>4</sub>, respectively. Given what we know about their inheritance patterns, their genotypes should be *Le*<sub>3</sub>*Le*<sub>3</sub>*le*<sub>4</sub>*le*<sub>4</sub> in *G. hirsutum*, and *le*<sub>3</sub>*le*<sub>3</sub>*Le*<sub>4</sub>*Le*<sub>4</sub> in R4-4. Data from this study supported previous information in that expression of the lethal symptom might be affected by the dosage of causal alleles and the environment in which plants are growing.

## Introduction

One of the classic definitions of a species is reproductive isolation, which is usually enforced by some physical, biochemical or behavioral mechanism. These restrictive

mechanisms can be classified into two groups: prezygotic and postzygotic. Hybrid lethality occurs because of postzygotic mechanisms that maintain species integrity and reproductive isolation (Stebbins 1966). Phenotypic manifestations of hybrid lethality include cell death, tissue necrosis, wilting, yellowing, reddening, chlorosis, dwarfism and reduced growth rate. Often these manifestations lead to actual death, although in a small number of cases the individual hybrids do reach reproductive maturity. Few, however, are sufficiently vigorous to reproduce. This phenomenon has been observed in F<sub>1</sub> hybrids, as well as later generations. Hybrid lethality, and the implications for speciation, have been described in the interspecific and intraspecific hybrids of several animals (Provine 1991; Sasa et al. 1998; Presgraves 2002; Price and Bouvier 2002) and plants (Hollingshead 1930; Wiebe 1934; Caldwell and Compton 1943; Savant 1956; Saunders 1952; McNaughton and Harper 1960; Tsunewaki 1970; Chu and Oka 1972; Sato and Hayashi 1983; Christie and MacNair 1984; Singh and Gutiérrez 1984; Valkonen and Watanabe 1999; Yamada et al. 2001; Mino et al. 2002; Moyle and Graham 2005; Marubashi and Tezuka 2006; Bomblyes et al. 2007).

Several cases of hybrid lethality in *Gossypium* (cotton) have been reported. Hutchinson (1932) studied crosses between certain strains of *G. arboreum* var. *soudanensis* and a range of other *arboreum* and *herbaceum* strains. Silow (1941) reported hybrid lethality in *G. herbaceum* and other species including *G. arboreum* and *G. anomalum*. Hybrid lethality has also been recognized between certain strains of *G. hirsutum* var. *marie-galante* and *G. barbadense* (Stephens 1946), Asiatic *G. arboreum* (sanguineum) and upland strains of *G. hirsutum* (Gerstel 1954), *G. hirsutum* and *G. gossypoides* (Philips and Merritt 1972), *G. klotzchianum* (D<sub>3-k</sub>) and other species containing A (two species), E (four species), F (one species) and other D genomes (Phillips and

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Reid 1975). In these cases, the hybrid phenotype resulted from the interaction of alleles at one or two loci in the parental genomes.

In the instance of *G. davidsonii*, Lee (1981a) suggested that the hybrid lethality resulted from allelic and interlocus interactions of  $Le_2^{dav}$  from *G. davidsonii* with  $Le_1$  and  $Le_2$  from *G. hirsutum* and *G. barbadense*.  $Le_1$  and  $Le_2$  were not linked to each other, but  $Le_2^{daw}$  was identified as a  $Le_2$  allele. Monotelodisomic tests indicated the  $Le_2$  locus was located in the long arm of chromosome D12, or within 5 cM of the centromere in the short arm (Stelly 1990). The  $Le_1$  locus was found in the long arm of chromosome A12; its distance from the centromere was about 13 cM (Samora et al. 1994).

Clarification of reproductive isolating mechanisms will provide important information about biological speciation and genes that may be employed for targeted crop breeding. To this end, representative examples are identified to study the underlying genetic basis and, subsequently, to clone the genes involved in the process. In this study, we identified hybrid lethality between *G. hirsutum* and *G. barbadense* cv. Coastland R4-4 (hereafter referred to as R4-4). We conducted genetic analyses and mapped the causative genes using a detailed linkage map (Guo et al. 2007).

## Materials and methods

### Plant materials

The *G. barbadense* cv. Coastland R4-4 (R4-4) line and three inbred lines of *G. hirsutum* were used, including a dominant fuzzless seed mutant ( $N_1FLM$ ), a recessive fuzzless seed mutant ( $n_2FLM$ ) and the genetic standard upland stock, TM-1. The phenotypes and origins of the parents are presented in Table 1.

The three inbred lines of *G. hirsutum*,  $N_1FLM$ ,  $n_2FLM$  and TM-1, were used as female parents in crosses with R4-4 at the Jiangpu Breeding Station of Nanjing Agricultural University (JBS-NAU), Jiangsu province of China. The  $F_1$

plants were self-pollinated to produce the  $F_2$ s and backcrossed to develop the  $BC_1$  on Hainan Island in China. In addition, TM-1 and  $N_1FLM$ , were used as male parents in crosses with R4-4 at JBS-NAU.

All the resulting  $F_1$ ,  $F_2$ , and  $BC_1$  plants as well as the parents were grown for field evaluations at JBS-NAU, and the numbers of plants are listed in Table 2. All these plants were grown in rows spaced 80 cm apart by row and 30 cm by plant. Field conditions consistent with those for normal cotton production were employed. Field evaluations for ( $N_1FLM \times$  Coastland R4-4) $F_2$ , ( $N_1FLM \times$  Coastland R4-4) $N_1FLM$ , ( $n_2FLM \times$  Coastland R4-4) $F_2$ , ( $n_2FLM \times$  Coastland R4-4) $n_2FLM$ , ( $N_1FLM \times$  Coastland R4-4) $F_1$  and ( $n_2FLM \times$  Coastland R4-4) $F_1$  along with their parents were conducted in 2006; other crosses as well as the parents were evaluated in 2007.

### Phenotypic analysis

The phenotype was scored every 2 weeks beginning at the first true leaf stage and continuing to maturity of individual plants. Phenotypes were designated as lethal or normal, depending upon leaf appearance. Leaves of unusual appearance characterized by reddening and lethal necrosis on the surface and back, were assigned to the lethal phenotype. Normal green leaves were marked as the normal phenotype.

For all of the field tests,  $F_1$  and the two parents were used as controls. The numbers of lethal and normal plants were counted in the  $F_2$  and  $BC_1$  populations. Segregation of lethal and normal plants was tested using a chi-square test to determine if the observed ratios fit Mendelian genetic models at the 5% level. Heterogeneity was assessed with chi-square to test if the segregation ratios are the same across different populations within  $F_2$  and  $BC_1$  progenies. The heterogeneity  $\chi^2$  value was obtained by subtracting the  $\chi^2$  value for the total number of plants from the sum of all the individual  $\chi^2$  values within  $F_2$  or  $BC_1$  populations, respectively. The heterogeneity degree of freedom was the number of crosses minus one within  $F_2$  or  $BC_1$  populations.

**Table 1** Phenotypes and origins of the parents

Materials	Phenotypes	Genotypes	Species	Origins
Texas Marker 1, TM-1	Normal	$Le_3Le_3le_4le_4$	<i>G. hirsutum</i>	USDA-ARS, College Station, Texas, USA
Dominant fuzzless mutant, $N_1FLM$	Normal	$Le_3Le_3le_4le_4$	<i>G. hirsutum</i>	USDA-ARS, College Station, Texas, USA
Recessive fuzzless mutant, $n_2FLM$	Normal	$Le_3Le_3le_4le_4$	<i>G. hirsutum</i>	Cotton Research Institute, Nanjing Agricultural University (CRI/NAU)
Coastland R4-4, R4-4	Normal	$le_3le_3Le_4Le_4$	<i>G. barbadense</i>	Coastal Plain Experiment Station, Tifton, Georgia, USA

**Table 2** Segregation ratios for sensitivity to hybrid lethality

Population/cross	No. lethal plants	No. normal plants	$\chi^2$	Probability %
( $N_1$ FLM $\times$ R4-4) $F_1$	105	0	–	–
( $n_2$ FLM $\times$ R4-4) $F_1$	38	0	–	–
(TM-1 $\times$ R4-4) $F_1$	32	0	–	–
(R4-4 $\times$ $N_1$ FLM) $F_1$	42	0	–	–
(R4-4 $\times$ TM-1) $F_1$	29	0	–	–
( $N_1$ FLM $\times$ R4-4) $F_2$	485	404	1.0370(9:7)	0.250–0.500
( $n_2$ FLM $\times$ R4-4) $F_2$	229	165	0.5610(9:7)	0.250–0.500
(TM-1 $\times$ R4-4) $F_2$	133	91	0.8889(9:7)	0.250–0.500
Total	847	660	0.0013(9:7)	>0.900
Heterogeneity ( $df = 2$ )			2.4856	0.250–0.500
( $N_1$ FLM $\times$ R4-4) $N_1$ FLM	160	183	1.5423(1:1)	0.100–0.250
( $N_1$ FLM $\times$ R4-4)R4-4	170	179	0.2321(1:1)	0.500–0.750
( $n_2$ FLM $\times$ R4-4) $n_2$ FLM	132	107	2.6151(1:1)	0.100–0.250
(TM-1 $\times$ R4-4)R4-4	181	224	4.5654(1:1)	0.025–0.050
(TM-1 $\times$ R4-4)TM-1	207	248	3.6945(1:1)	0.050–0.100
Total	850	941	4.6237(1:1)	0.025–0.050
Heterogeneity ( $df = 4$ )			8.0257	0.050–0.100

#### DNA extraction, PCR amplification, and electrophoresis

Cotton genomic DNA extraction followed the procedure of Paterson et al. (1993). Simple sequence repeat polymerase chain reaction (SSR-PCR) amplifications were performed using a Peltier Thermal Cycler-225 (MJ Research, USA). Electrophoresis of the products was conducted as described by Zhang et al. (2000, 2002).

#### Molecular mapping of the hybrid lethality genes

In comparison with the  $F_2$  population,  $BC_1$  populations are able to generate more genotypic information for linkage analysis since only one of the two necrosis genes actually segregates, making it easier to identify genotype. The two backcross populations ( $N_1$ FLM  $\times$  R4-4)R4-4 and ( $N_1$ FLM  $\times$  R4-4) $N_1$ FLM, consisting of 349 and 343 individuals, respectively, were used to map the causal genes.

For the linkage maps, normal and lethal plants were scored as 1 and 3, respectively, and the parental genotype and the heterozygous genotype were scored as 1 and 3, respectively, in the two  $BC_1$  populations. Missing data were designated as “–”. The chi-square ( $\chi^2$ ) test was used to determine if normal segregation had occurred at the 5% level.

JoinMap 3.0 (Van Ooijen and Voorrips 2001) was employed to construct the genetic linkage map. Recombination frequency was converted to genetic map distance (centiMorgen, cM) using the Kosambi mapping function (Kosambi 1944). The linkage groups were isolated with log-of-odds (LOD) scores  $\geq 10$ .

#### Results

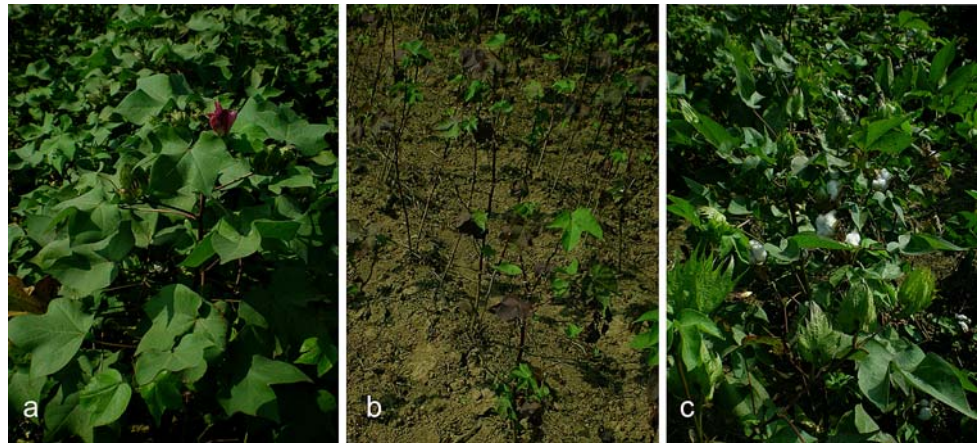
##### Hybrid lethality in interspecific hybrids between *G. hirsutum* and R4-4

To map the fuzzless genes  $N_1$  and  $n_2$  in cotton, the two fuzzless seed mutants  $N_1$ FLM and  $n_2$ FLM, containing  $N_1$  and  $n_2$ , respectively, were crossed with R4-4 with fuzzy seed. However, the lethal phenotype was detected in these  $F_1$  hybrids. To further study the lethal symptom, a genetic standard line of Upland cotton, TM-1, was used for an additional cross with R4-4.

In the interspecific crosses between *G. hirsutum* acc.  $N_1$ FLM,  $n_2$ FLM and TM-1 with R4-4 (Table 2), all  $F_1$  seeds initially germinated and developed normally. However, the lethal symptoms appeared in the  $F_1$  plants during the pre-budding stage, beginning with the first true leaf. While the young leaves initially appeared normal, they quickly developed a reddish color between veins on both the top and back surfaces. Leaf color gradually changed to purple and the leaves began drooping. Finally, the mature leaves dropped, leaving naked stems behind (Fig. 1b).

The  $F_1$  hybrids grown in Nanjing developed more severe lethal symptoms, and produced no flowers. However, the  $F_1$  hybrid plants from Hainan Island (in southern China) were normal, and produced viable seed, allowing for additional research. The observed variability in the degree of lethal symptoms may be attributable to environmental conditions, as has been previously reported (Gerstel 1954; Philips 1977; Shii et al. 1980; Manabe et al. 1989; Yamada et al. 2000; Yamada and Marubashi 2003; Bombliet et al. 2007).

**Fig. 1** Phenotype of *N<sub>7</sub>FLM*, *N<sub>7</sub>FLM/R4-4* F<sub>1</sub> hybrid, and R4-4 at the open boll stage; **a–c** denote *N<sub>7</sub>FLM*, *N<sub>7</sub>FLM/F<sub>1</sub>* hybrids and R4-4, respectively



#### Genetic analysis of the lethal phenotype in the interspecific hybrids

To investigate the genetic mechanism(s) producing the lethal phenotype, three inbred lines, *N<sub>7</sub>FLM*, *n<sub>2</sub>FLM* and TM-1 in *G. hirsutum* and R4-4, along with their resulting F<sub>1</sub>, F<sub>2</sub> and BC<sub>1</sub> progenies, were grown at JBS-NAU. The parents were normal, but in the interspecific crosses of *N<sub>7</sub>FLM* × R4-4, *n<sub>2</sub>FLM* × R4-4 and TM-1 × R4-4, all F<sub>1</sub> hybrids showed similar symptoms, indicating that the lethal characteristic was caused by an interaction of genes from *G. hirsutum* and R4-4 (Fig. 1). Their reciprocal F<sub>1</sub> hybrids, R4-4 × *N<sub>7</sub>FLM* and R4-4 × TM-1, exhibited the same lethal phenotype as their respective original cross, indicating that the cytoplasmic genome had no impact on the symptoms.

The segregations of (*N<sub>7</sub>FLM* × R4-4)F<sub>2</sub>, (*n<sub>2</sub>FLM* × R4-4)F<sub>2</sub> and (TM-1 × R4-4)F<sub>2</sub> fit a two dominant complementary genes model, with a 9:7 (lethal:normal) ratio. Likewise, the BC<sub>1</sub> population of (*N<sub>7</sub>FLM* × R4-4)*N<sub>7</sub>FLM*, (*n<sub>2</sub>FLM* × R4-4)*n<sub>2</sub>FLM*, (TM-1 × R4-4)TM-1 and (*N<sub>7</sub>FLM* × R4-4)R4-4 segregated in a 1:1 (lethal:normal) ratio, as expected under the two dominant complementary genes model. There was a slight deviation ( $\chi^2 = 4.5654$ ) in (TM-1 × R4-4)R4-4. Homogeneity tests indicated that populations within F<sub>2</sub> and BC<sub>1</sub> progenies were homogeneous (Table 2).

In these F<sub>2</sub> and backcross populations, some plants had more severe symptoms than the F<sub>1</sub> hybrids, and died at an early developmental stage, including the seedling stage (Fig. 2). The earlier the symptoms of lethality were manifested in the plants, the more severe they were. Symptoms occurred earlier in F<sub>2</sub> than in backcross populations. The variation of lethal symptoms within a population (Fig. 2) was probably caused by the dosage of causal alleles, as previously reported (Shii et al. 1980; Rooney and Stelly 1990; Chu et al. 2006; Bomblies et al. 2007; Ichitani et al. 2007).

All of these data indicate that the lethal phenotype in interspecific hybrids is controlled by two dominant

complementary genes: one derived from the *G. hirsutum* parent and the other from R4-4 in *G. barbadense*. We did not know then if the genes identified in the present research are the same as *Le<sub>1</sub>* and *Le<sub>2</sub>* that were reported by Lee (1981a, b). Therefore, they were tentatively designated as *Le<sub>h</sub>* from *G. hirsutum* and *Le<sub>b</sub>* from *G. barbadense* cv. Coastland R4-4.

#### Molecular mapping of two dominant complementary genes responsible for the hybrid lethality

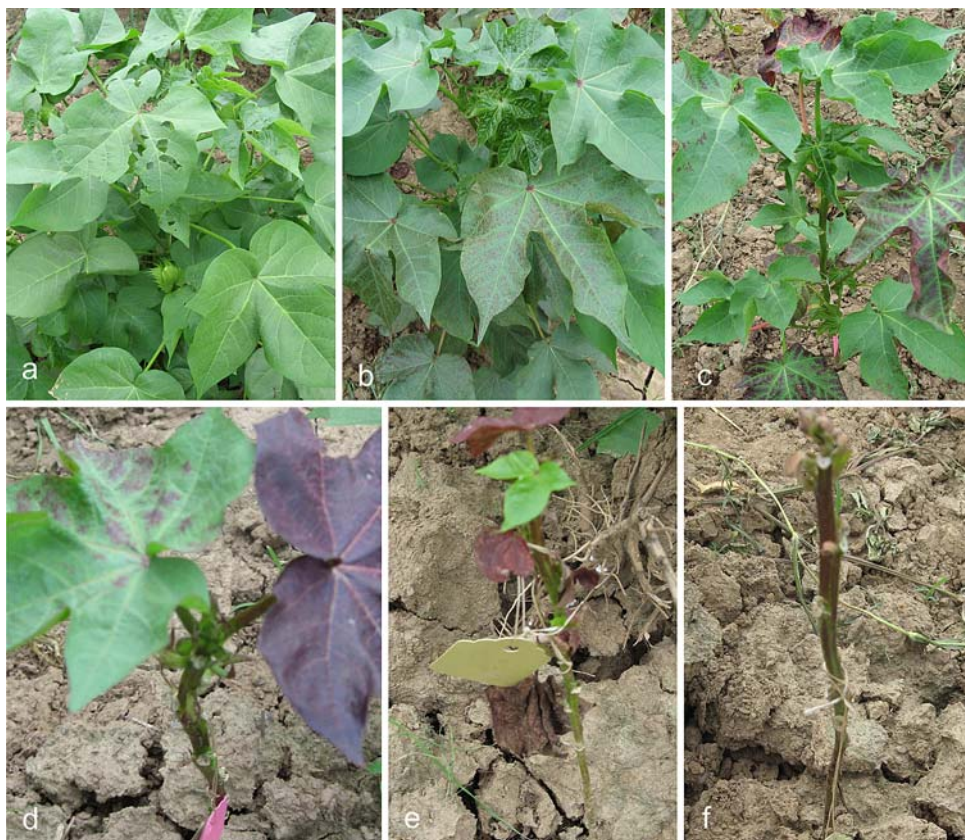
Because the lethal phenotype in interspecific hybrids is controlled by two dominant complementary genes, it is possible to segregate each one separately in backcross (BC<sub>1</sub>) populations. To map *Le<sub>h</sub>* from *G. hirsutum*, R4-4 was used as the recurrent parent to produce the backcross population, whereas, to map *Le<sub>b</sub>* from *G. barbadense*, *G. hirsutum* cultivars were used as the recurrent parents.

#### Molecular mapping of *Le<sub>h</sub>* derived from *G. hirsutum*

The (*N<sub>7</sub>FLM* × R4-4) R4-4 backcross population, comprised of 349 individuals, was used to map the hybrid lethality gene *Le<sub>h</sub>* from *G. hirsutum*. Earlier studies determined that the two genes, *Le<sub>1</sub>* and *Le<sub>2</sub>*, interacted with an allele at the *Le<sub>2</sub>* locus–*Le<sub>2</sub><sup>daw</sup>* to induce hybrid lethality; *Le<sub>1</sub>* and *Le<sub>2</sub>* were assigned to A12 (Chr. 12) and D12 (Chr. 26), respectively (Lee 1981b; Stelly 1990; Samora et al. 1994). Given this location information, SSR markers anchored on Chr. A12 and Chr. D12 (Guo et al. 2007) at about 10 cM intervals were used to screen 65 randomly selected plants from the (*N<sub>7</sub>FLM* × R4-4) R4-4 population. Linkage analysis by JoinMap 3.0, however, failed to detect any marker on Chr. A12 and Chr. D12 that was linked to the hybrid lethality gene *Le<sub>h</sub>*.

In order to quickly determine on which chromosome the *Le<sub>h</sub>* gene was found, the bulked segregant analysis (BSA) method (Michelmore et al. 1991) was employed. Seven plants with lethal characteristics and seven normal plants

**Fig. 2** Observed phenotypes in the  $F_2$  population of the cross  $N_7FLM \times R4-4$  at the budding stage. **a** Wild type (wt); **b–f** lethal types

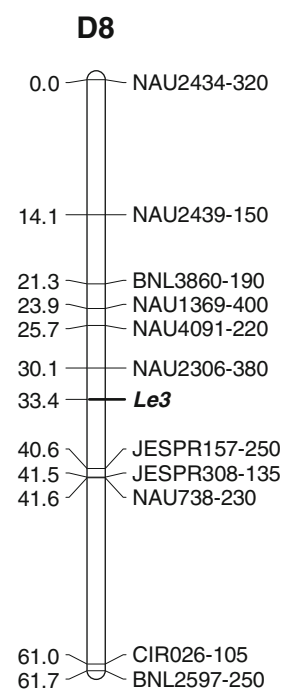


from the mapping population were selected to build the lethal and normal pools, respectively. Four hundred and thirty (430) SSR markers, positioned at about 10 cM intervals, were selected from all chromosomes (24) except A12 and D12 (Guo et al. 2007) to detect the polymorphism between the parents and DNA pools. The analysis identified three SSR markers—NAU2306, NAU738 and NAU2439—on Chr. D8 that were polymorphic between the two parents and two pools, suggesting that these three SSR markers may be linked to the hybrid lethality gene  $Le_h$ . The three SSR markers were further used to screen their association with the  $Le_h$  gene using the same 65 randomly selected individuals from the  $(N_7FLM \times R4-4)R4-4$  population. Linkage analysis indicated that  $Le_h$  was linked to these same three SSR markers on Chr. D8. Eleven of the SSR markers around  $Le_h$  from the linkage group in Chr. D8 (Guo et al. 2007) were then selected to analyze the whole population for fine mapping of the gene  $Le_h$ . The resulting map illustrated the position of  $Le_h$  relative to 11 neighboring markers on Chr. D8 (Fig. 3). The closest marker to  $Le_h$  was NAU2306, at a distance of 3.3 cM.

#### Molecular mapping of $Le_b$ from R4-4

The  $(N_7FLM \times R4-4)N_7FLM$  population, comprised of 343 individuals, was used to map  $Le_b$ , the complementary

**Fig. 3** Linkage map of the region of D8 surrounding  $Le_3$  derived from *G. hirsutum* from analysis of the  $(N_7FLM \times R4-4)R4-4$  population



hybrid lethality gene from *G. barbadense*. Because  $Le_h$  was mapped on Chr. D8, another hybrid lethality gene,  $Le_y$ , was expected to be on Chr. A8, a homoeologous chromosome of Chr. D8. SSR markers anchored on Chr. A8 (Guo et al.

2007) at about 10 cM intervals were used to initially screen 93 randomly selected plants from the ( $N_7$ FLM  $\times$  R4-4)  $N_7$ FLM population. Linkage analysis, however, failed to associate any marker on Chr. A8 with  $Le_b$ .

Using the BSA method (Michelmore et al. 1991), seven lethal plants and seven normal plants from the mapping populations were selected to build the lethal and normal pools, respectively. 455 SSR markers, drawn from all 25 chromosomes other than A8 (Guo et al. 2007), were selected at about 10 cM intervals to screen for polymorphism. Four markers, NAU3740, BNL3279, NAU2602 and NAU3889 anchored on Chr. D11, were found to be polymorphic between the two parents and between the two pools, suggesting that the  $Le_b$  gene might lie on Chr. D11. The four polymorphic markers were further used to screen their association with the  $Le_b$  gene with the same 93 randomly selected individuals from the backcross population ( $N_7$ FLM  $\times$  R4-4)  $N_7$ FLM. As expected, the gene  $Le_b$  was linked to the four SSR markers on Chr. D11. Nine of the SSR markers located around  $Le_b$  from the linkage group in Chr. D11 (Guo et al. 2007) were selected to screen the 343 individuals for fine mapping of the gene  $Le_b$ . The resulting map illustrated the position of  $Le_b$  relative to nine neighboring markers on D11 (Fig. 4). The nearest marker to  $Le_b$  was BNL1154, at a distance of 4.1 cM.

The mapping of  $Le_h$  in *G. hirsutum* on D8 and  $Le_b$  in R4-4 on D11 indicates that these genes differ from  $Le_1$  and  $Le_2$ , which are located on A12 and D12, respectively. Therefore, they are novel loci responsible for hybrid lethality in *Gossypium*. Based on standard genetic nomenclature (Kohel 1973),  $Le_h$  from *G. hirsutum* and  $Le_b$

from R-4 were tentatively designated as  $Le_3$  and  $Le_4$ , respectively. The corresponding genotypes are  $Le_3Le_3le_4le_4$  in *G. hirsutum*, and  $le_3le_3Le_4Le_4$  in *G. barbadense* cv. R4-4.

## Discussion

### Hybrid lethality in *Gossypium*

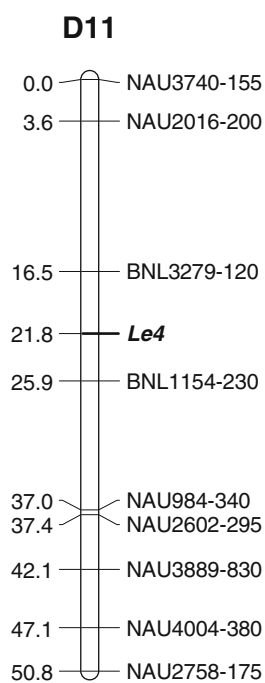
Hybrid lethality plays an important role in reproductive isolation and speciation (Orr and Presgraves 2000). To date, several cases of interspecific hybrids exhibiting lethality have been reported in cotton (Stephens 1946, Phillips 1976, Lee 1981a). Stephens (1946) reported the presence of lethal hybrids between certain strains of *G. hirsutum* var. *marie-galante* and *G. barbadense*. The hybrids have a characteristic bushy habit owing to their shortened internodes and excess production of lateral branches; leaves are inrolled and exhibit a yellowish mottling. The symptoms depend on the interaction of two dominant complementary alleles located on the same chromosome in the D-genome, carried by the *marie-galante* and the *G. barbadense* parents. Phillips (1976) characterized the lethal symptom as tumorigenesis. In addition, Lee (1981a) identified the lethal symptom in interspecific hybrids of *G. davidsonii* Kell. (D<sub>3</sub>) and cultivated tetraploid cottons, *G. hirsutum* and *G. barbadense*. Lethality resulted from the intralocus and interlocus interaction of  $Le_2^{dav}$  (the D<sub>3</sub> complementary hybrid lethality system factor) with alleles at the  $Le_1$  and/or  $Le_2$  loci (Lee 1981a, 1981b). Linkage and monotelodisomic tests indicated the  $Le_2$  locus was located in chromosome D12, and  $Le_1$  in chromosome A12 (Stelly 1990; Samora et al. 1994; Wang et al. 2006).

In this study, hybrid lethality between *G. hirsutum* and R4-4 was characterized by leaf reddening and necrosis, leaving bare stems and leading to death in many cases. Through genetic analysis we were able to determine that the deleterious characteristics were controlled by two dominant complementary genes: one on D8 in *G. hirsutum* and the other on D11 in R4-4. Based upon a review of similar research in cotton, the interactions of these two genes appear to be a novel mechanism for the induction of hybrid lethality in these two taxa.

Dobzhansky–Muller genes act as an interspecific isolating mechanism between *G. hirsutum* and *G. barbadense* cv. Coastland R4-4

In our study,  $Le_3$  and  $Le_4$ , the two dominant complementary genes from *G. hirsutum* and R4-4, interact to cause lethality in interspecific hybrids, a situation which supports the Dobzhansky–Muller model, where hybrid incompatibilities,

**Fig. 4** Linkage map of the region of D11 surrounding  $Le_4$  derived from *G. barbadense* from analysis of the ( $N_7$ FLM  $\times$  R4-4)  $N_7$ FLM population



such as death or sterility, are caused by genes that have evolved from a common ancestor but diverged in each of the species. In the common ancestor, these genes may have worked in a complementary manner, but evolutionary changes have resulted in their current antagonism (Dobzhansky 1937; Muller 1942; Brideau et al. 2006; Bomblies and Weigel 2007).

Based on the Dobzhansky–Muller model, we suggest that *G. hirsutum* and R4-4 alleles evolved from a common ancestor with a genotype of  $le_3le_3le_4le_4$ . Through evolution,  $le_3$  diverged in *G. hirsutum* to  $Le_3$ , and  $le_4$  diverged in R4-4 to  $Le_4$ . While functioning well within the genome of each species, the  $Le_3$ – $Le_4$  hybrid interaction is detrimental. This hypothesis is consistent with previous propositions wherein *G. hirsutum* and *G. barbadense* diverged from the original AD tetraploid entity, and the AD tetraploid species ( $2n = 4x = 52$ ) arose from interspecific hybridization between A- and D-genome diploid *Gossypium* species (Seelanan et al. 1997; Small et al. 1998).

Although *G. barbadense* and *G. hirsutum* are identified as two distinct species, most of the interspecific hybrids between these species are vigorous, and some hybrids are used as commercial varieties. A species can be defined as a group of actually, or potentially, interbreeding natural populations that are reproductively isolated from other such groups (Mayr 1942). In this study, we discovered one case of interspecific hybrid lethality between *G. hirsutum* and *G. barbadense* cv. Coastland R4-4, thus demonstrating the existence of mechanisms to discourage hybridization between them. Our data indicate the Dobzhansky–Muller genes,  $Le_3$  and  $Le_4$ , act as an interspecific isolating mechanism between *G. hirsutum* and *G. barbadense* cv. Coastland R4-4.

*Gossypium barbadense* originated in South America (Hutchinson 1959). Until the 16th century, it was the primary fiber crop on the continent (Kerr 1960). The ancestral cotton plants were photoperiodic, and the fiber was medium staple (23.8–27.0 mm in length) and coarse, as typified by current Tanguis cottons of Peru. *Gossypium barbadense* first appeared in the United States ca. 1785, where it became known as Sea Island (Hutchinson 1959). Sea Island is characterized as long staple fiber. The exact origin of Sea Island is unknown, but Kerr (1960) and Stephens (1975, 1976) suggested that the most logical development path was by transgressive inheritance through the introgression of length genes from outside the species, possibly from *G. hirsutum*. Coastland, on the other hand, was developed from intraspecific crossing of three *G. barbadense* types: Sea Island, Egyptian and Tanguis (Jenkins 1953). We presume, therefore, that the hybrid lethality allele in the other cultivars of *G. barbadense* was eliminated or modified by selection, whereas in R4-4 it was maintained by crossing of the three *G. barbadense* types. If this is the case, the lethal

combination evaluated in our study might be novel and provide additional insight into the divergence of *G. hirsutum* and *G. barbadense*.

In the present paper, the two novel Dobzhansky–Muller genes,  $Le_3$  and  $Le_4$ , were both mapped on D-genome chromosomes. In the Dobzhansky–Muller model, the hybrid lethality genes are differentiated between the parent lineages. Evidence from several studies of *Drosophila* and *A. thaliana* suggest that hybrid lethality may arise as a by-product of adaptive evolution, since the few known Dobzhansky–Muller genes are all rapidly evolving (Coyne and Orr 2004; Orr 2005; Mallet 2006; Bomblies et al. 2007). The evolutionary rates and levels of nucleotide diversity are positively correlated (Hudson et al. 1987). We suppose that the differentiated hybrid lethality genes between *G. hirsutum* and R4-4 might be a result of differential evolutionary pressure acting on the two sub-genomes. This hypothesis is strongly supported by previous reports that indicate the D-subgenome harbors greater nucleotide and allelic diversity than the A-subgenome in both *G. hirsutum* and *G. barbadense*, based on a comparison of duplicated paralogous *Adh* loci (Small and Wendel 2002; Small et al. 1999). However, an understanding of the evolution of  $Le_3$  and  $Le_4$  must await their cloning and identification of their biochemical function. Their fine mapping will help to clone them.

#### Molecular mechanisms of hybrid lethality

By clarifying the mechanism(s) associated with hybrid lethality, important insights into biological speciation and the general properties of postzygotic reproductive isolation are obtained. As an initial step to clone the causal genes following the Dobzhansky–Muller model, several genes have been positively located on linkage maps for many plant taxa, including wheat, rice and *Arabidopsis* (Chu et al. 2006; Ichitani et al. 2001, 2007; Bomblies et al. 2007). Some of the Dobzhansky–Muller genes in animals that have been cloned are the *Drosophila* Hybrid male rescue (*Hmr*) and its interacting partner Lethal hybrid rescue (*Lhr*) (Barbash et al. 2003; Brideau et al. 2006), and *Nucleoporin96* (*Nup96*) (Presgraves et al. 2003; Presgraves and Stephan 2007). Diverse functions are indicated for these genes. It has been speculated that the MYB-related gene *Hmr*, which is expressed at all stages during the life cycle, plays a role in normal gene regulation (Barbash et al. 2003). *Lhr* is an interacting partner of Heterochromatin Protein 1 (HP1) with, as yet, an unknown function (Brideau et al. 2006). *Nup96* encodes a structural protein of the nuclear pore complex (Presgraves et al. 2003). Common features of these genes are their rapid evolution (Coyne and Orr 2004; Orr 2005; Mallet 2006) and positive Darwinian selection (Barbash et al. 2003; Presgraves et al. 2003; Orr 2005;

Presgraves and Stephan 2007). In *Arabidopsis*, one allele of an *NB-LRR* gene interacts with another allele at the same locus to cause hybrid lethality (Bomblies et al. 2007). The *NB-LRR* genes are the most common type of plant disease-resistance genes. The known diversity of these genes has been interpreted as the result of positive selection (Bakker et al. 2006). If this is the case, however, it suggests the possibility that selective pressure exerted by pathogens can promote rapid evolution of gene variants that may prove beneficial to the parents, but deleterious to hybrid progeny. This link between hybrid lethality and disease-resistance pathways has also been suggested in tobacco (Masuda et al. 2007). Bomblies et al. (2007) and Bomblies and Weigel (2007) hypothesized that hybrid lethality could result from autoimmunity, perhaps as a pleiotropic effect of the evolution of genes that are involved in pathogen response.

Based upon the extant literature, it is apparent that studies of the molecular mechanisms underlying hybrid lethality have primarily focused on the model organisms, *Drosophila* sp. and *Arabidopsis thaliana*. Because the hybrid lethality mechanisms are potentially diverse, additional examples can only help to strengthen our understanding of the evolution of postzygotic reproductive isolation and speciation. In cotton, however, no genes causing hybrid lethality have been tagged with molecular markers, or cloned. The current study is the first occasion where two Dobzhansky–Muller genes have been located on the genetic linkage maps with SSR markers. The next step toward a more complete understanding of the key molecular mechanisms must be cloning, which will further uncover the structure, function, products and actions of the two causal genes.

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## References

- Bakker EG, Toomajian C, Kreitman M, Bergelson J (2006) A genome-wide survey of R gene polymorphisms in *Arabidopsis*. *Plant Cell* 18:1803–1818
- Barbash DA, Siino DF, Tarone AM, Roote J (2003) A rapidly evolving MYB related protein causes species isolation in *Drosophila*. *Proc Natl Acad Sci USA* 100:5302–5307
- Bomblies K, Weigel D (2007) Hybrid necrosis: autoimmunity as a common barrier to gene flow in plants. *Nat Rev Genet* 8:382–393
- Bomblies K, Lempe J, Epple P, Warthmann N, Lanz C, Dangl JL, Weigel D (2007) Autoimmune response as a mechanism for a Dobzhansky–Muller-type incompatibility syndrome in plants. *PLoS Biol* 5(9):1962–1972
- Brideau NJ, Flores HA, Wang J, Maheshwari S, Wang X, Barbash DA (2006) Two Dobzhansky–Muller genes interact to cause hybrid lethality in *Drosophila*. *Science* 314:1292–1295
- Caldwell RM, Compton LE (1943) Complementary lethal genes in wheat causing a progressive lethal necrosis of seedlings. *J Hered* 34:67–70
- Christie P, MacNair MR (1984) Complementary lethal factors in two North American populations of the yellow monkey flower. *J Hered* 75:510–511
- Chu Y, Oka H (1972) The distribution and effects of genes causing F1 weakness in *Oryza breviligulata* and *O. glaberrima*. *Genetics* 70:163–173
- Chu CG, Faris JD, Friesen TL, Xu SS (2006) Molecular mapping of hybrid necrosis genes *Ne1* and *Ne2* in hexaploid wheat using microsatellite markers. *Theor Appl Genet* 112:1374–1381
- Coyne JA, Orr HA (2004) Speciation. Sinauer Associates, Inc, Sunderland, MA
- Dobzhansky TH (1937) Genetics and the origin of species. Columbia University Press, New York
- Gerstel DU (1954) A new lethal combination in interspecific cotton hybrids. *Genetics* 39:628–639
- Guo WZ, Cai CP, Wang CB, Han ZG, Song XL, Wang K, Niu XW, Wang C, Lu KY, Shi B, Zhang TZ (2007) A microsatellite-based, gene-rich linkage map reveals genome structure, function, and evolution in *Gossypium*. *Genetics* 176:527–541
- Hollingshead LA (1930) A lethal factor in *Crepis* effective only in an interspecific hybrid. *Genetics* 15:114–140
- Hudson RR, Kreitman MK, Aguade M (1987) A test of neutral molecular evolution based on nucleotide data. *Genetics* 116:153–159
- Hutchinson JB (1932) The genetics of cotton. VII. ‘Crumpled’, a new dominant in Asiatic cottons produced by complementary factors. *J Genet* 25:281–291
- Hutchinson J (1959) The application of genetics to cotton improvement. Cambridge University Press, London
- Ichitani K, Fukuta Y, Taura S, Sato M (2001) Chromosomal location of *Hwc2*, one of the complementary hybrid weakness genes, in rice. *Plant Breed* 120:523–525
- Ichitani K, Namigoshi K, Sato M, Taura S, Aoki M (2007) Fine mapping and allelic dosage effect of *Hwc1*, a complementary hybrid weakness gene in rice. *Theor Appl Genet* 114:1407–1415
- Jenkins JG (1953) Coastland-A new long staple cotton for the Southeast. Georgia Coastal Plain Exp Stn Bull 53
- Kerr T (1960) The potentials of barbadense cottons. In: Proceedings of the 12th cotton improvement conference, Memphis, pp 57–60
- Kohel RJ (1973) Genetic nomenclature in cotton. *Hered* 64:291–295
- Kosambi DD (1944) The estimation of map distance from recombination values. *Ann Eugen* 12:172–175
- Lee JA (1981a) Genetics of D3 complementary lethality in *Gossypium hirsutum* and *G. barbadense*. *J Hered* 72:299–300
- Lee JA (1981b) A new linkage relationship in cotton. *Crop Sci* 21:346–347
- Mallet J (2006) What does *Drosophila* genetics tell us about speciation? *Trends Ecol Evol* 21:386–393
- Manabe T, Marubashi W, Onozawa Y (1989) Temperature-dependent conditional lethality in interspecific hybrids between *Nicotiana suaveolens* Lehm. and *N. tabacum* L. In: Proceedings of the 6th International Congress of the Society for the Advancement of Breeding Researches in Asia and Oceania (SABRAO), Philippine Rice Research Institute, Philippines. pp 459–462
- Marubashi W, Tezuka T (2006) Hybrid lethality in interspecific hybrids between *Nicotiana tabacum* and *N. suaveolens*: evidence that the Q chromosome causes hybrid lethality based on Q-chromosome-specific DNA markers. *Theor Appl Genet* 112:1172–1178
- Masuda Y, Yamada T, Kuboyama T, Marubashi W (2007) Identification and characterization of genes involved in hybrid lethality in hybrid tobacco cells (*Nicotiana suaveolens* × *N. tabacum*) using suppression subtractive hybridization. *Plant Cell Rep* 26:1595–1604
- Mayr E (1942) Systematics and the origin of species. Columbia University Press, New York
- McNaughton IH, Harper JL (1960) The comparative biology of closely related species living in the same area. II. Aberrant

- morphology and a virus-like syndrome in hybrids between *Papaver rhoeas* L. and *P. dubium* L. New Phytol 59:27–41
- Michelmore RW, Paran IP, Kesseli RV (1991) Identification of markers linked to disease resistance genes by bulked segregant analysis: a rapid method to detect markers in specific genomic regions by using segregating populations. Proc Natl Acad Sci USA 88:9828–9832
- Mino M, Maekawa K, Ogawa K, Yamagishi H, Inoue M (2002) Cell death processes during expression of hybrid lethality in interspecific F<sub>1</sub> hybrid between *Nicotiana glauca* Domin and *Nicotiana glauca*. Plant Physiol 130:1776–1787
- Moyle LC, Graham EB (2005) Genetics of hybrid incompatibility between *Lycopersicon esculentum* and *L. hirsutum*. Genetics 169:355–373
- Muller HJ (1942) Isolating mechanisms, evolution, and temperature. Biol Symp 6:71–125
- Orr HA (2005) The genetic basis of reproductive isolation: insights from *Drosophila*. Proc Natl Acad Sci USA 102(1):6522–6526
- Orr HA, Presgraves DC (2000) Speciation by postzygotic isolation: forces, genes and molecules. BioEssays 22:1085–1094
- Paterson AH, Brubaker C, Wendel JF (1993) A rapid method for extraction of cotton (*Gossypium* spp.) genomic DNA suitable for RFLP or PCR analysis. Plant Mol Biol Rep 11:122–127
- Philips LL (1977) Interspecific incompatibility in *Gossypium*. IV. Temperature-conditional lethality in hybrids of *G. klotzschianum*. Am J Bot 64:914–915
- Philips LL, Merritt JF (1972) Interspecific compatibility in *Gossypium*. I. Stem histogenesis of *G. hirsutum* × *G. gossypoides*. Am J Bot 59:203–208
- Phillips LL (1976) Interspecific incompatibility in *Gossypium*. III. The genetics of tumorigenesis in hybrids of *G. gossypoides*. Can J Genet Cytol 18:365–369
- Phillips LL, Reid RK (1975) Interspecific compatibility in *Gossypium*. II. Light and electron microscopic studies of cell necrosis and tumorigenesis in hybrid of *G. klotzschianum*. Am J Bot 62:790–796
- Presgraves DC (2002) Patterns of postzygotic isolation in *Lepidoptera*. Evolution 56:1168–1183
- Presgraves DC, Stephan W (2007) Pervasive adaptive evolution among interactors of the *Drosophila* hybrid inviability gene, *Nup96*. Mol Biol Evol 24:306–314
- Presgraves DC, Balagopalan L, Abmayr SM, Orr HA (2003) Adaptive evolution drives divergence of a hybrid inviability gene between two species of *Drosophila*. Nature 423:715–719
- Price TD, Bouvier MM (2002) The evolution of F<sub>1</sub> postzygotic incompatibilities in birds. Evolution 56:2083–2089
- Provine WB (1991) Alfred Henry Sturtevant and crosses between *Drosophila melanogaster* and *Drosophila simulans*. Genetics 129:1–5
- Rooney WL, Stelly DM (1990) Genetic effects on the timing of *Le2<sup>daw</sup>* induced necrosis of cotton. Crop Sci 30:70–74
- Samora PJ, Stelly DM, Kohel RJ (1994) Localization and mapping of the *Le1* and *Gl2* loci of cotton (*Gossypium hirsutum* L.). J Hered 85:152–156
- Sasa MM, Chippendale PT, Johnson NA (1998) Patterns of postzygotic isolation in frogs. Evolution 52:1811–1820
- Sato YI, Hayashi J (1983) Distribution of the complementary genes causing F<sub>1</sub> weakness in the common rice and its wild relatives I. *L-2-a* gene in Asian native cultivars. Jpn J Genet 58:411–418
- Saunders AR (1952) Complementary lethal genes in the cowpea. S Afr J Sci 48:195–197
- Savant AC (1956) Semilethal complementary factors in a tomato species hybrid. Evolution 10:93–96
- Seelanan T, Schnabel A, Wendel JF (1997) Congruence and consensus in the cotton tribe. Syst Bot 22:259–290
- Shii CT, Mok MC, Temple SR, Mok DWS (1980) Expression of developmental abnormalities of *Phaseolus vulgaris* L. Interaction between temperature and allelic dosage. J Hered 71:219–222
- Silow RA (1941) The comparative genetics of *Gossypium anomalum* and the cultivated Asiatic cottons. J Genet 42:259–358
- Singh SP, Gutiérrez JA (1984) Geographical distribution of the *DL1* and *DL2* genes causing hybrid dwarfism in *Phaseolus vulgaris* L., their association with seed size, and their significance to breeding. Euphytica 33:337–345
- Small RL, Wendel JF (2002) Differential evolutionary dynamics of duplicated paralogous *Adh* loci in allotetraploid cotton (*Gossypium*). Mol Biol Evol 19:597–607
- Small RL, Ryburn JA, Cronn RC, Seelanan T, Wendel JF (1998) The tortoise and the hare: choosing between noncoding plastome and nuclear *Adh* sequences for phylogeny reconstruction in a recently diverged plant group. Am J Bot 85:1301–1315
- Small RL, Ryburn JA, Wendel JF (1999) Low levels of nucleotide diversity at homoeologous *Adh* loci in allotetraploid cotton (*Gossypium* L.). Mol Biol Evol 16:491–501
- Stebbins GL (1966) Reproductive isolation and the origin of species. In: Processes of organic evolution. Prentice-Hall, New Jersey. pp 85–112
- Stelly DM (1990) Localization of the *Le2* Locus of cotton (*Gossypium hirsutum* L.). J Hered 81:193–197
- Stephens SG (1946) The genetics of *Corky*. Part I. The new world alleles and their possible role as an interspecific isolating mechanism. J Genet 47:150–161
- Stephens SG (1975) Some observations on photoperiodism and the development of annual forms of domesticated cottons. Econ Bot 30:409–418
- Stephens SG (1976) The origin of Sea Island cotton. Agric History 50:391–399
- Tsunewaki K (1970) Necrosis and chlorosis genes in common wheat and its ancestral species. Seiken Zihō 22:67–75
- Valkonen JPT, Watanabe KN (1999) Autonomous cell death, temperature sensitivity and the genetic control associated with resistance to cucumber mosaic virus (CMV) in diploid potatoes (*Solanum* spp.). Theor Appl Genet 99:996–1005
- Van Ooijen JW, Voorrips RE (2001) JoinMapR Version 3.0: software for the calculation of genetic linkage maps. Centre for Plant Breeding and Reproduction Research (CPRO-DLO). Wageningen, The Netherlands
- Wang K, Song XL, Han ZG, Guo WZ, Yu JZ, Sun J, Pan JJ, Kohel RJ, Zhang TZ (2006) Complete assignment of the chromosomes of *Gossypium hirsutum* L. by translocation and fluorescence in situ hybridization mapping. Theor Appl Genet 113:73–80
- Wiebe GA (1934) Complementary factors in barley giving a lethal progeny. J Hered 25:272–274
- Yamada T, Marubashi W (2003) Overproduced ethylene causes programmed cell death leading to temperature-sensitive lethality in hybrid seedlings from the cross *Nicotiana suaveolens* × *N. tabacum*. Planta 217:690–698
- Yamada T, Marubashi W, Niwa M (2000) Apoptotic cell death induces temperature-sensitive lethality in hybrid seedlings and calli derived from the cross of *Nicotiana suaveolens* × *N. tabacum*. Planta 211:614–622
- Yamada T, Marubashi W, Niwa M (2001) Facile induction of apoptosis into plant cells associated with temperature-sensitive lethality shown on interspecific hybrid from the cross *Nicotiana suaveolens* × *N. tabacum*. Plant Cell Physiol 42:204–213
- Zhang J, Wu YT, Guo WZ, Zhang TZ (2000) Fast screening of microsatellite markers in cotton with PAGE/silver staining. Cotton Sci 12:267–269
- Zhang J, Guo WZ, Zhang TZ (2002) Molecular linkage map of allotetraploid cotton (*Gossypium hirsutum* L. × *Gossypium barbadense* L.) with a haploid population. Theor Appl Genet 105:1166–1174